

## Synthesis of (2*E*)-3-{2-[(substituted benzyl)oxy]phenyl}acrylaldehydes as novel anti-inflammatory agents

Li-Jiau Huang,<sup>a,\*</sup> Jih-Pyang Wang,<sup>a,b,\*</sup> Yu-Chi Lai<sup>a</sup> and Sheng-Chu Kuo<sup>a</sup>

<sup>a</sup>Graduate Institute of Pharmaceutical Chemistry, China Medical University, Taichung 40402, Taiwan, ROC

<sup>b</sup>Department of Education and Research, Taichung Veterans General Hospital, Taichung 407, Taiwan, ROC

Received 26 September 2005; revised 28 January 2006; accepted 6 February 2006

Available online 28 February 2006

**Abstract**—As part of our continuing effort for development of novel anti-inflammatory agents, the highly potential agent **CCY1a**, which we reported recently, was selected as lead compound to synthesize a series of its derivatives for evaluation. Most of the newly synthesized compounds exhibited superior inhibitory activity than both the lead compound and the positive control (trifluoperazine) toward fMLP-stimulated neutrophil superoxide formation. (2*E*)-3-[2-(Benzlyoxy)-5-methoxyphenyl]-acrylaldehyde (**31**) was among the most potent with action mechanism different from **CCY1a** in that it does not act as cAMP-elevating agent but inhibits the increase in cellular Ca<sup>2+</sup> with greater potency.

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In our previous work,<sup>1</sup> several regioisomers of benzyl-oxybenzaldehyde were synthesized and determined to have significant inhibitory activity toward neutrophil superoxide anion generation. Among these compounds, 2-benzyloxybenzaldehyde (**1**, **CCY1a**) was the most promising one, which was further identified as an indirect adenylyl cyclase activator,<sup>2</sup> through the release of endogenous adenosine. The inhibition of superoxide anion generation by **CCY1a** in neutrophils is attributed partly to its cAMP-elevating activity, and partly to the blockade of external Ca<sup>2+</sup> entry and phospholipase D activation via cAMP-independent mechanisms.<sup>2–4</sup>

In the present work, **CCY1a** was selected as lead compound and a series of 3-{2-[(substituted benzyl)oxy]phenyl}acrylaldehydes were synthesized and their effect on neutrophil superoxide formation and cAMP-elevating activity were evaluated.

The (2*E*)-3-{2-[(substituted benzyl)oxy]phenyl}acrylaldehydes (**15–26**) were synthesized according to the synthetic method of **CCY1a**.<sup>1</sup> As shown in [Scheme 1](#),

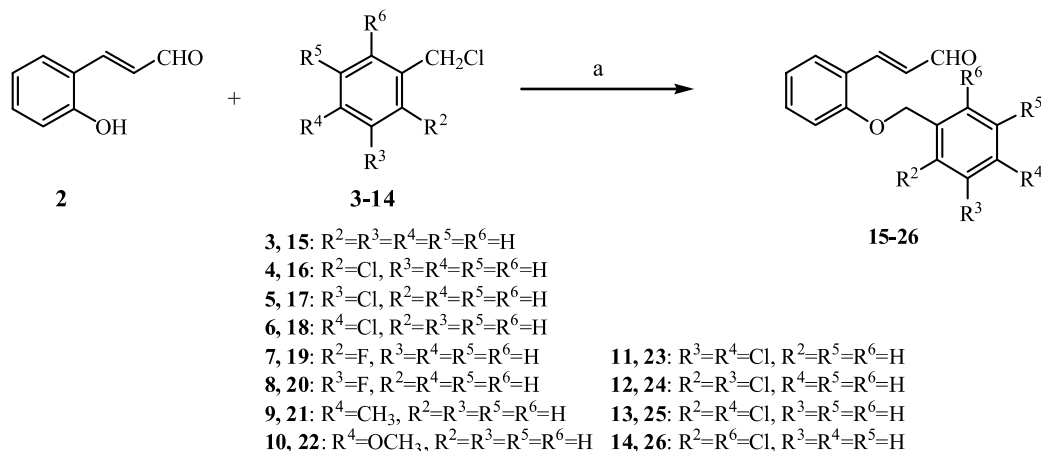
when cinnamic aldehyde (**2**) was reacted with a variety of substituted benzyl chlorides (**3–14**) and K<sub>2</sub>CO<sub>3</sub> in EtOH, the corresponding target compounds (**15–26**) were obtained. On the other hand, as illustrated in [Scheme 2](#), 2-(substituted benzyloxy)-5-methoxybenzaldehyde (**27–30**),<sup>5</sup> which was prepared from 2-hydroxy-5-methoxybenzaldehyde and substituted benzyl chloride (**3–6**), was treated with equal mole of acetaldehyde at low temperature to undergo aldol condensation which resulted in the formation of (2*E*)-3-[2-(substituted benzyloxy)-5-methoxyphenyl]acrylaldehyde (**31–34**). Finally, compounds **31–33** were treated with hydroxylamine to afford the corresponding oximes **35–37**. All the synthetic compounds were characterized by their spectral characteristics.<sup>6</sup>

Superoxide anion formation was measured in terms of superoxide dismutase inhibitable cytochrome *c* reduction.<sup>7</sup> Briefly, the assay mixture contained the cell suspension and 0.5 mg/mL of cytochrome *c* ( $\Delta A_{550}$ ), the reference mixture also received 6.6 µg/mL of superoxide dismutase. Reaction was initiated by stimulating with 0.3 µM formyl-Met-Leu-Phe (fMLP) or 3 nM phorbol 12-myristate 13-acetate (PMA).

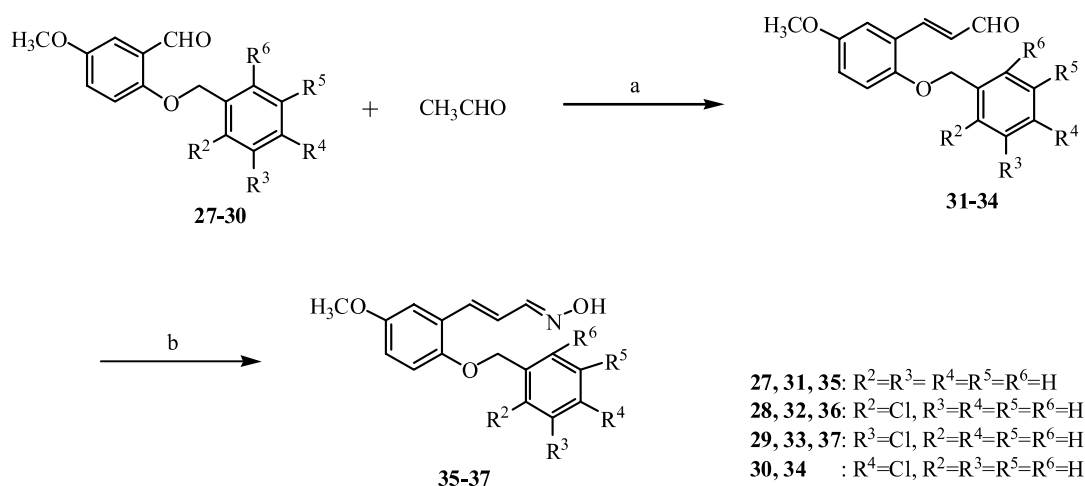
According to the data in [Table 1](#), converting the –CHO moiety of lead compound (**CCY1a**) into –CH=CH–CHO (**15**) improved its activity significantly. Moreover, the addition of a Cl atom into the *ortho*

**Keywords:** Anti-inflammatory agents; 3-{2-[(Substituted benzyl)-oxy]phenyl}-acrylaldehydes.

\* Corresponding authors. Tel.: +886 4 22053366 1007; fax: +886 04 22055105 (L.J.H.); tel.: +886 04 23592525 2043; fax: +886 04 23592705 (J.P.W.); e-mail addresses: [ljhuang@mail.cmu.edu.tw](mailto:ljhuang@mail.cmu.edu.tw); [w1994@vghtc.vghtc.gov.tw](mailto:w1994@vghtc.vghtc.gov.tw)



**Scheme 1.** Reagents and conditions: (a)  $K_2CO_3$ , KI/ethanol, reflux.



**Scheme 2.** Reagents and conditions: (a) 20% NaOH/ethanol, 0 °C; (b)  $NH_2OH \cdot HCl$ ,  $CH_3COONa$ /ethanol, reflux.

position of the benzyl group in compound **15** resulted in compound **16** with further improved activity. Meanwhile, the placement of the same Cl atom in the *meta* position (**17**) appeared to be functionless, whereas a Cl atom in *para* position (**18**) resulted in reduced activity. In order approaches, it was found that the substitution of the *ortho* Cl of compound **16**, or the *meta* Cl of **17** with F atom, resulted in compounds **19** and **20** with a slight improvement in their activity. Then our attempt to modify compound **15** by adding a  $CH_3$  group onto its *para* position at benzyl group resulted in compound **21** with only about half the activity.

Results on the effect of all dichlorobenzyl derivatives (**23–26**) on neutrophil superoxide anion formation indicated that only 3,4-dichlorobenzyl derivatives (**23**) demonstrated significant activity. Neither its 2,3-dichloro (**24**), 2,4-dichloro (**25**) nor 2,6-dichloro (**26**) isomers show any significant activity. Excitingly, the incorporation of an  $OCH_3$  group into the 4 position at the benzyl-oxy group (**22**) and the 5 position at the phenyl group (**31**) of compound **15** resulted in significant boost of activity.

In particular, the  $IC_{50}$  value for compound **31** was determined as 2.6  $\mu M$  which is about 6 times more active than the lead compound **CCY1a** or the positive control (trifluoperazine). After introduction of a Cl atom onto the benzyl group of compound **31** at either the *ortho*- (**32**), *meta*- (**33**) or *para*-location (**34**), only the 3-chlorobenzyl derivatives (**33**) maintain significant inhibitory activity against superoxide formation. Our attempt in derivatization of compound **31** into an oxime (**35**) resulted in 50% lower activity, whereas similar derivatization of compound **32** resulted in compound **36** with improved activity in 30  $\mu M$  concentration. On the other hand, all the compounds were tested for their inhibitory activity against PMA-stimulated superoxide anion formation. The result indicated that, unlike the positive control TFP, none of the tested compounds showed significant activity in the test concentration range 1–30  $\mu M$ . This seems to suggest that, the action mechanism of the test compounds differs when tested against superoxide anion generation by different stimulants.

Summarizing all the above findings, compound **31** was found to demonstrate the greatest inhibitory activity against fMLP-induced superoxide anion formation of

**Table 1.** The inhibitory effects of **15–26** and **31–37** on the neutrophil superoxide formation

				Inducer: fMLP (0.3 $\mu$ M)/cytochalasin B (5 $\mu$ g/mL)			
Animal: Rat (Sprague Dawley)				Inducer: PMA (3 nM)			
Compound	R'	Concentration ( $\mu$ M)	Superoxide formation (nmol/ $10^6$ cells/30 min)				
			fMLP	% inhibition	PMA	% inhibition	
Control			2.89 $\pm$ 0.30		7.60 $\pm$ 0.36		
CCY1a	C <sub>6</sub> H <sub>5</sub>	3	1.01 $\pm$ 0.19**	36.1 $\pm$ 6.3	—	—	
		10	0.86 $\pm$ 0.20**	40.6 $\pm$ 5.9	—	—	
		30	0.62 $\pm$ 0.07**	60.8 $\pm$ 1.6	—	—	
			IC <sub>50</sub> = 15.6				
<b>15</b>	C <sub>6</sub> H <sub>5</sub>	3	1.96 $\pm$ 0.19	31.8 $\pm$ 3.3**	—	—	
		10	1.05 $\pm$ 0.16	63.7 $\pm$ 2.4**	8.61 $\pm$ 0.50	–13.5 $\pm$ 6.0	
		30	0.48 $\pm$ 0.05	83.2 $\pm$ 1.4**	9.39 $\pm$ 0.49	–23.6 $\pm$ 4.8	
			IC <sub>50</sub> = 6.4 $\pm$ 0.7				
<b>16</b>	2-Cl-C <sub>6</sub> H <sub>4</sub>	3	1.69 $\pm$ 0.13	40.1 $\pm$ 7.5**	—	—	
		10	0.82 $\pm$ 0.08	71.4 $\pm$ 2.3**	9.86 $\pm$ 0.27	–30.1 $\pm$ 5.4	
		30	0.29 $\pm$ 0.02	89.7 $\pm$ 0.7**	8.39 $\pm$ 0.36	–10.9 $\pm$ 7.0	
			IC <sub>50</sub> = 4.4 $\pm$ 1.1				
<b>17</b>	3-Cl-C <sub>6</sub> H <sub>4</sub>	3	2.00 $\pm$ 0.26	31.2 $\pm$ 1.8**	—	—	
		10	0.9 $\pm$ 0.12	65.7 $\pm$ 2.6**	9.85 $\pm$ 0.14	–30.1 $\pm$ 5.6	
		30	0.34 $\pm$ 0.04	88.0 $\pm$ 1.5**	9.06 $\pm$ 0.89	–18.7 $\pm$ 7.8	
			IC <sub>50</sub> = 5.9 $\pm$ 0.3				
<b>18</b>	4-Cl-C <sub>6</sub> H <sub>4</sub>	3	2.23 $\pm$ 0.20	22.4 $\pm$ 1.3	—	—	
		10	1.05 $\pm$ 0.06	63.1 $\pm$ 2.2**	9.41 $\pm$ 0.35	–23.9 $\pm$ 3.1	
		30	0.37 $\pm$ 0.02	86.9 $\pm$ 1.3**	9.26 $\pm$ 0.31	–22.0 $\pm$ 3.7	
			IC <sub>50</sub> = 7.4 $\pm$ 0.3				
<b>19</b>	2-F-C <sub>6</sub> H <sub>4</sub>	3	1.48 $\pm$ 0.12	47.5 $\pm$ 7.0**	—	—	
		10	0.72 $\pm$ 0.13	75.1 $\pm$ 2.3**	9.45 $\pm$ 0.64	–24.1 $\pm$ 4.3	
		30	0.35 $\pm$ 0.05	87.6 $\pm$ 2.2**	9.82 $\pm$ 0.51	–29.4 $\pm$ 6.0	
			IC <sub>50</sub> = 3.3 $\pm$ 0.8				
<b>20</b>	3-F-C <sub>6</sub> H <sub>4</sub>	3	1.70 $\pm$ 0.12	40.3 $\pm$ 3.6**	—	—	
		10	0.76 $\pm$ 0.06	73.0 $\pm$ 3.4**	8.07 $\pm$ 0.26	–6.2 $\pm$ 2.3	
		30	0.46 $\pm$ 0.06	83.9 $\pm$ 0.9**	9.38 $\pm$ 0.39	–23.7 $\pm$ 6.5	
			IC <sub>50</sub> = 4.5 $\pm$ 0.7				
<b>21</b>	4-CH <sub>3</sub> -C <sub>6</sub> H <sub>4</sub>	3	2.24 $\pm$ 0.15	21.9 $\pm$ 2.8	—	—	
		10	1.57 $\pm$ 0.11	45.1 $\pm$ 1.9**	8.53 $\pm$ 0.48	–13.2 $\pm$ 11.0	
		30	0.78 $\pm$ 0.12	73.2 $\pm$ 1.2**	9.51 $\pm$ 0.18	–25.8 $\pm$ 7.7	
			IC <sub>50</sub> = 11.0 $\pm$ 0.4				
<b>22</b>	4-OCH <sub>3</sub> -C <sub>6</sub> H <sub>4</sub>	3	1.58 $\pm$ 0.09	44.6 $\pm$ 2.8**	—	—	
		10	0.81 $\pm$ 0.02	71.3 $\pm$ 1.8**	9.45 $\pm$ 0.51	–24.5 $\pm$ 6.3	
		30	0.33 $\pm$ 0.09	88.2 $\pm$ 3.7**	9.28 $\pm$ 0.31	–22.2 $\pm$ 1.9	
			IC <sub>50</sub> = 3.8 $\pm$ 0.5				
<b>23</b>	3,4-Cl-C <sub>6</sub> H <sub>3</sub>	1	2.56 $\pm$ 0.35	11.7 $\pm$ 3.1	—	—	
		3	2.03 $\pm$ 0.21	29.4 $\pm$ 2.4*	—	—	
		10	0.60 $\pm$ 0.20	79.8 $\pm$ 4.4**	7.77 $\pm$ 0.29	1.9 $\pm$ 6.3	
		30	—	—	11.4 $\pm$ 0.48	–51.1 $\pm$ 8.6*	
			IC <sub>50</sub> = 4.3 $\pm$ 0.4				
<b>24</b>	2,3-Cl-C <sub>6</sub> H <sub>3</sub>	10	2.39 $\pm$ 0.27	17.4 $\pm$ 0.8	6.72 $\pm$ 0.39	11.2 $\pm$ 7.2	
		30	1.75 $\pm$ 0.30	39.9 $\pm$ 4.9**	6.29 $\pm$ 0.38	17.3 $\pm$ 1.2	
<b>25</b>	2,4-Cl-C <sub>6</sub> H <sub>3</sub>	10	2.16 $\pm$ 0.17	24.4 $\pm$ 4.7	9.29 $\pm$ 0.06	–22.7 $\pm$ 6.6	
		30	1.76 $\pm$ 0.33	39.9 $\pm$ 5.5**	9.84 $\pm$ 0.57	–29.8 $\pm$ 8.4	
<b>26</b>	2,6-Cl-C <sub>6</sub> H <sub>3</sub>	10	1.71 $\pm$ 0.21	40.3 $\pm$ 7.5**	11.4 $\pm$ 0.55	–50.6 $\pm$ 5.3*	
		30	1.54 $\pm$ 0.27	46.8 $\pm$ 7.6**	12.0 $\pm$ 0.14	–59.2 $\pm$ 7.6**	
<b>31</b>	C <sub>6</sub> H <sub>5</sub>	1	2.09 $\pm$ 0.10	26.7 $\pm$ 4.1*	—	—	
		3	1.37 $\pm$ 0.16	52.24 $\pm$ 2.4**	—	—	
		10	0.53 $\pm$ 0.03	81.2 $\pm$ 1.0**	9.42 $\pm$ 0.52	–24.0 $\pm$ 5.4	
		30	—	—	9.19 $\pm$ 0.37	–20.9 $\pm$ 2.8	
			IC <sub>50</sub> = 2.6 $\pm$ 0.2				

Table 1 (continued)

Compound	R'	Concentration (μM)	Superoxide formation (nmol/10 <sup>6</sup> cells/30 min)			
			fMLP	% inhibition	PMA	% inhibition
32	2-Cl-C <sub>6</sub> H <sub>4</sub>	10	1.89 ± 0.18	34.3 ± 2.6**	8.94 ± 0.59	−17.5 ± 3.7
		30	1.69 ± 0.13	40.9 ± 1.9**	9.63 ± 0.39	−26.9 ± 5.4
33	3-Cl-C <sub>6</sub> H <sub>4</sub>	1	2.54 ± 0.17	11.4 ± 3.4	—	—
		3	1.82 ± 0.20	36.8 ± 1.4**	—	—
		10	0.67 ± 0.22	77.7 ± 5.6**	10.3 ± 0.91	−35.7 ± 8.4
		30	—	—	9.96 ± 0.67	−31.2 ± 7.7
IC <sub>50</sub> = 4.2 ± 0.3						
34	4-Cl-C <sub>6</sub> H <sub>4</sub>	10	2.10 ± 0.21	27.1 ± 0.5*	9.39 ± 0.72	−23.3 ± 6.0
		30	1.90 ± 0.08	33.4 ± 4.3**	9.55 ± 0.66	−25.5 ± 5.8
35	C <sub>6</sub> H <sub>5</sub>	3	2.03 ± 0.08	41.9 ± 3.4**	—	—
		10	1.05 ± 0.08	70.0 ± 1.3**	4.97 ± 0.88	7.6 ± 12.5
		30	0.48 ± 0.05	86.0 ± 1.9**	6.90 ± 0.76	−29.4 ± 8.3
IC <sub>50</sub> = 4.3 ± 0.5						
36	2-Cl-C <sub>6</sub> H <sub>4</sub>	3	2.45 ± 0.30	29.5 ± 10.5*	—	—
		10	2.09 ± 0.23	40.4 ± 5.8**	5.27 ± 0.64	1.5 ± 7.7
		30	0.78 ± 0.11	77.7 ± 2.9**	7.62 ± 0.12	−44.4 ± 4.7
IC <sub>50</sub> = 9.5 ± 1.4						
37	4-Cl-C <sub>6</sub> H <sub>4</sub>	10	2.53 ± 0.22	27.9 ± 4.6*	11.9 ± 0.25	−124 ± 13.7**
		30	2.41 ± 1.4	31.6 ± 7.9**	12.1 ± 0.15	−132 ± 14.4**
TFP		1	—	—	5.48 ± 0.13	27.6 ± 4.2*
		3	28.0 ± 0.27	1.7 ± 6.6	3.4 ± 0.13	55.4 ± 9.4**
		5	2.1 ± 0.09	24.2 ± 5.3*	—	—
		10	0.54 ± 0.22	79.5 ± 4.2**	1.23 ± 0.16	83.3 ± 5.8**
			IC <sub>50</sub> = 6.6 ± 0.2		IC <sub>50</sub> = 2.7 ± 0.6	

—, not determined. Trifluoperazine (TFP): positive control.

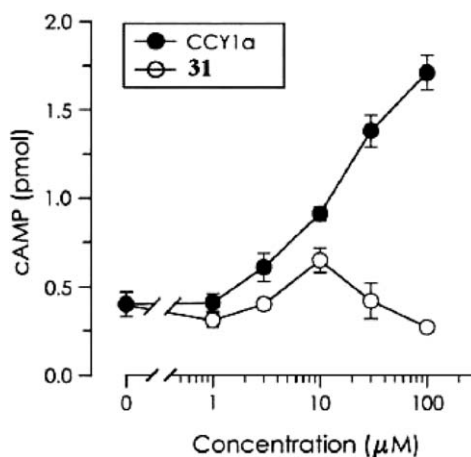
\*  $P < 0.05$ .

\*\*  $P < 0.01$ ;  $N = 3$ .

neutrophil. Therefore, compound **31** was selected for study of action mechanism.

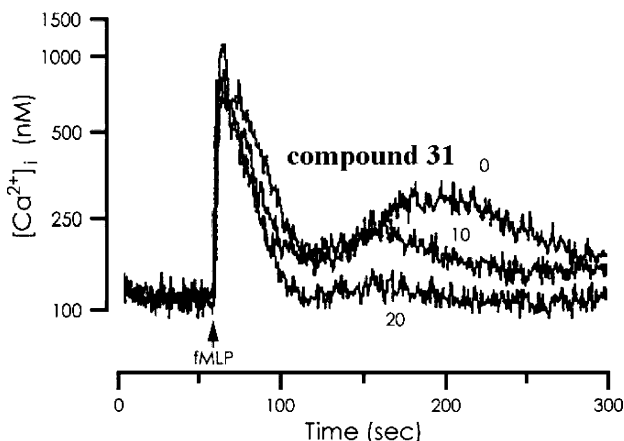
We also determined the cAMP-elevating activity of compound **31** in neutrophils by using an enzyme immunoassay kit (Amersham) as the previous report.<sup>1</sup> Surprisingly, compound **31** only slightly elevated the cellular cAMP level at 10  $\mu\text{M}$  and then returned to the baseline at higher concentrations (Fig. 1), unlike **CCY1a**, which produced a concentration-dependent increase in cellular cAMP level. We next determined the effect of compound **31** on intracellular free  $\text{Ca}^{2+}$  concentration because the fMLP-induced superoxide anion is a  $\text{Ca}^{2+}$ -dependent process and **CCY1a** also exerts its effect through the inhibition of  $\text{Ca}^{2+}$  entry.<sup>2</sup> In this experiment, neutrophils were loaded with 5  $\mu\text{M}$  fluo-3 AM for 45 min. After being washed, the cells were resuspended and monitored for the fluorescence change (488/535 nm). Addition of fMLP to fluo-3-loaded cells evoked an initial  $\text{Ca}^{2+}$  spike, followed by a plateau phase, in the presence of 1 mM extracellular  $\text{Ca}^{2+}$ .<sup>8</sup> Interestingly, compound **31** was 2.5-fold more potent than **CCY1a** in the inhibition of plateau phase (Fig. 2).

To sum up, the inhibitory activity for all of the above **CCY1a** derivatives against neutrophil superoxide formation was found to be far superior than



**Figure 1.** Effect of **CCY1a** and compound **31** on cellular cAMP levels. Cells were treated with DMSO (as basal level), the indicated concentrations of **CCY1a** or compound **31** for 10 min. The cAMP contents were assayed using an enzyme immunoassay kit. Values are means  $\pm$  SD of three experiments.

both their lead compound and the positive control. The action of the most potent compound **31** is attributed at least partly to the blockade of intracellular  $\text{Ca}^{2+}$  elevation but not to the cAMP-dependent mechanism.



**Figure 2.** Effect of compound **31** on the fMLP-stimulated elevation of intracellular free  $\text{Ca}^{2+}$  in neutrophils. Fluo 3-loaded cells in HBSS containing 1 mM  $\text{Ca}^{2+}$  were pretreated with the indicated concentrations of compound **31** at 37 °C for 1 min before stimulation with 0.3  $\mu\text{M}$  fMLP. The data presented are representative of three independent experiments with similar results.

### Acknowledgments

This work was supported by research grants from the National Science Council of the Republic of China (NSC91-2320-B-039-039) and in part by China Medical University (CMC90-PC-03) awarded to L. J. Huang.

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- Spectral characteristics for compounds: Compound **15**: yield, 42%;  $^1\text{H}$  NMR (200 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  9.63 (d,  $J$  = 7.8 Hz, 1H), 7.90 (d,  $J$  = 16.0 Hz, 1H), 7.73 (d,  $J$  = 7.5 Hz, 1H), 7.29–7.50 (m, 6H), 7.19 (d,  $J$  = 8.4 Hz, 1H), 7.02 (dd,  $J$  = 7.5, 7.5 Hz, 1H), 6.75 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.23 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  194.80, 157.02, 147.79, 136.56, 133.26, 132.80, 129.35, 129.03, 128.54, 127.98, 127.65, 122.68, 121.04, 69.81 ppm; MS (EI)  $m/z$  238 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{14}\text{O}_2$ : C, 80.65; H, 5.92. Found: C, 79.98; H, 5.86. Compound **16**: yield, 35%;  $^1\text{H}$  NMR (200 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  9.62 (d,  $J$  = 7.8 Hz, 1H), 7.89 (d,  $J$  = 16.0 Hz, 1H), 7.70 (d,  $J$  = 7.5, 1.2 Hz, 1H), 7.37–7.65 (m, 5H), 7.23 (d,  $J$  = 7.2 Hz, 1H), 7.05 (dd,  $J$  = 7.5, 7.5 Hz, 1H), 6.82 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.29 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  194.82, 156.78, 147.58, 133.80, 133.32, 132.90, 130.38, 129.54, 129.32, 129.12, 127.48, 121.38, 120.13, 67.62 ppm; MS (EI)  $m/z$  272 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{13}\text{O}_2\text{Cl}$ : C, 70.46; H, 4.80. Found: C, 70.32; H, 4.68. Compound **17**: yield, 31%;  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.62 (d,  $J$  = 7.8 Hz, 1H), 7.88 (d,  $J$  = 16.0 Hz, 1H), 7.77 (dd,  $J$  = 7.5, 1.6 Hz, 1H), 7.25–7.56 (m, 5H), 6.99 (d,  $J$  = 8.4 Hz, 1H), 6.96 (dd,  $J$  = 7.5, 7.5 Hz, 1H), 6.79 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.13 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{CDCl}_3$ ):  $\delta$  194.35, 156.93, 147.59, 138.31, 134.64, 132.62, 130.06, 129.13, 128.67, 128.40, 127.37, 125.31, 123.38, 121.45, 112.68, 69.71 ppm; MS (EI)  $m/z$  272 ( $\text{M}^+$ ); Anal. Calcd for: C, 70.46; H, 4.80. Found: C, 70.82; H, 5.04. Compound **18**: yield, 43%;  $^1\text{H}$  NMR (200 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  9.63 (d,  $J$  = 7.8 Hz, 1H), 7.90 (d,  $J$  = 16.0 Hz, 1H), 7.72 (d,  $J$  = 7.5 Hz, 1H), 7.38–7.52 (m, 5H), 7.16 (d,  $J$  = 8.4 Hz, 1H), 7.01 (dd,  $J$  = 7.5, 7.5 Hz, 1H), 6.83 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.18 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  194.73, 156.78, 147.63, 135.54, 132.76, 132.65, 129.46, 129.24, 129.01, 128.52, 122.69, 121.13, 113.19, 68.96 ppm; MS (EI)  $m/z$  272 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{13}\text{O}_2\text{Cl}$ : C, 70.46; H, 4.80. Found: C, 69.88; H, 4.56. Compound **19**: yield, 33%;  $^1\text{H}$  NMR (200 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  9.60 (d,  $J$  = 7.8 Hz, 1H), 7.85 (d,  $J$  = 16.0 Hz, 1H), 7.75 (dd,  $J$  = 7.6, 1.4 Hz, 1H), 7.59 (ddd,  $J$  = 7.6, 7.6, 1.4 Hz, 1H), 7.41–7.47 (m, 2H), 7.21–7.32 (m, 3H), 7.04 (dd,  $J$  = 7.6, 7.6 Hz, 1H), 6.81 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.28 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  195.17, 160.72, 157.10, 148.02, 133.15, 130.97, 130.89, 129.56, 124.85, 123.52, 122.97, 121.61, 115.73, 113.54, 64.56 ppm; MS (EI)  $m/z$  256 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{13}\text{O}_2\text{F}$ : C, 74.99; H, 5.11. Found: C, 73.56; H, 4.88. Compound **20**: yield, 34%;  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.70 (d,  $J$  = 7.8 Hz, 1H), 7.91 (d,  $J$  = 16.0 Hz, 1H), 7.06 (dd,  $J$  = 8.0, 1.6 Hz, 1H), 7.33–7.44 (m, 2H), 6.95–7.27 (m, 5H), 6.78 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.17 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{CDCl}_3$ ):  $\delta$  194.18, 156.78, 147.43, 138.54, 132.40, 130.15, 128.95, 128.49, 122.45, 122.31, 121.22, 114.95, 113.97, 112.47, 69.51 ppm; MS (EI)  $m/z$  256 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{13}\text{O}_2\text{F}$ : C, 74.99; H, 5.11. Found: C, 75.65; H, 4.89. Compound **21**: yield, 31%;  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.67 (d,  $J$  = 7.8 Hz, 1H), 7.89 (d,  $J$  = 16.0 Hz, 1H), 7.58 (dd,  $J$  = 8.0, 1.8 Hz, 1H), 7.20–7.43 (m, 4H), 6.97–7.05 (m, 2H), 6.77 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.13 (s, 2H), 2.38 (s, 3H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{CDCl}_3$ ):  $\delta$  194.52, 157.45, 147.41, 138.09, 133.21, 132.61, 129.41, 128.98, 128.66, 127.54, 123.36, 121.09, 112.82, 70.54, 21.20 ppm; MS (EI)  $m/z$  252 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{17}\text{H}_{16}\text{O}_2$ : C, 80.93; H, 6.39. Found: C, 79.88; H, 6.54. Compound **22**: yield, 32%;  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.66 (d,  $J$  = 7.8 Hz, 1H), 7.88 (d,  $J$  = 16.0 Hz, 1H), 7.58 (dd,  $J$  = 8.0, 1.8 Hz, 1H), 7.34–7.40 (m, 3H), 6.92–7.05 (m, 4H), 6.73 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.10 (s, 2H), 3.83 (s, 3H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{CDCl}_3$ ):  $\delta$  194.54, 159.66, 157.49, 148.08, 132.62, 123.39, 121.11, 114.15, 112.87, 70.44, 55.33 ppm; MS (EI)  $m/z$  268 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{17}\text{H}_{16}\text{O}_3$ : C, 76.10; H, 6.01. Found: C, 76.58; H, 5.98. Compound **23**: yield, 22%;  $^1\text{H}$  NMR (200 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  9.64 (d,  $J$  = 7.8 Hz, 1H), 7.92 (d,  $J$  = 16.0 Hz, 1H), 7.75–7.77 (m, 2H), 7.66 (d,  $J$  = 8.3 Hz, 1H), 7.41–7.50 (m, 2H), 7.17 (d,  $J$  = 8.4 Hz, 1H), 7.03 (dd,  $J$  = 7.5, 7.5 Hz, 1H), 6.83 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.25 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{DMSO}-d_6$ ):  $\delta$  194.92, 156.59, 147.65, 137.83, 132.89, 131.20, 130.84, 130.63, 129.68, 129.29, 129.13, 127.97, 122.76, 121.37, 113.30, 68.29 ppm; MS (EI)  $m/z$  306 ( $\text{M}^+$ ); Anal. Calcd for  $\text{C}_{16}\text{H}_{12}\text{O}_2\text{Cl}_2$ : C, 62.56; H, 3.94. Found: C, 63.51; H, 4.05. Compound **24**: yield, 24%;  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.71 (d,  $J$  = 7.8 Hz, 1H), 7.93 (d,  $J$  = 16.0 Hz, 1H), 7.61 (dd,  $J$  = 7.5, 1.4 Hz, 1H), 7.36–7.49 (m, 3H), 7.22–7.29 (m, 1H), 7.05 (d,  $J$  = 7.5, 7.5 Hz, 1H), 6.95 (d,  $J$  = 8.4 Hz, 1H), 6.78 (dd,  $J$  = 16.0, 7.8 Hz, 1H), 5.28 (s, 2H) ppm;  $^{13}\text{C}$  NMR (50 MHz,  $\text{CDCl}_3$ ):  $\delta$  194.31, 156.66, 147.50, 136.22, 133.31, 132.68, 130.75, 129.97, 129.10, 128.59, 127.56, 126.61, 123.35, 121.62, 112.72, 67.98 ppm; MS (EI)  $m/z$  306 ( $\text{M}^+$ ); Anal. Calcd

for  $C_{16}H_{12}O_2Cl_2$ : C, 62.56; H, 3.94. Found: C, 63.21; H, 4.01. Compound **25**: yield, 42%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.62 (d,  $J = 7.8$  Hz, 1H), 7.89 (d,  $J = 16.0$  Hz, 1H), 7.77 (d,  $J = 7.5$  Hz, 1H), 7.71 (d, 1H), 7.63 (m, 1H), 7.47–7.51 (m, 2H), 7.05 (d,  $J = 8.4$  Hz, 1H), 6.95 (dd,  $J = 7.5, 7.5$  Hz, 1H), 6.78 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.27 (s, 2H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.90, 156.63, 147.52, 133.87, 133.05, 132.95, 131.72, 129.29, 129.14, 127.70, 122.84, 121.53, 113.36, 67.06 ppm; MS (EI)  $m/z$  306 ( $M^+$ ); Anal. Calcd for  $C_{16}H_{12}O_2Cl_2$ : C, 62.56; H, 3.94. Found: C, 62.54; H, 3.88. Compound **26**: yield, 27%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.52 (d,  $J = 7.8$  Hz, 1H), 7.73 (d,  $J = 16.0$  Hz, 1H), 7.72 (d,  $J = 1.2$  Hz, 1H), 7.35–7.60 (m, 5H), 7.08 (dd,  $J = 7.5, 7.5$  Hz, 1H), 6.73 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.34 (s, 2H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.92, 157.35, 147.65, 136.32, 133.18, 132.00, 131.45, 129.74, 129.41, 129.12, 123.14, 121.94, 113.83, 66.11 ppm; MS (EI)  $m/z$  306 ( $M^+$ ); Anal. Calcd for  $C_{16}H_{12}O_2Cl_2$ : C, 62.56; H, 3.94. Found: C, 61.22; H, 4.02. Compound **28**: yield, 79%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  10.30 (s, 1H), 7.26–7.38 (m, 4H), 7.25 (d,  $J = 2.8$  Hz, 1H), 7.15 (d,  $J = 2.8$  Hz, 1H), 5.23 (s, 2H), 3.72 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  189.14, 155.13, 153.78, 134.00, 132.87, 130.31, 129.67, 127.64, 125.36, 123.24, 116.53, 110.57, 68.69, 55.75 ppm; MS (EI)  $m/z$  276 ( $M^+$ ); Anal. Calcd for  $C_{15}H_{13}O_3Cl$ : C, 65.11; H, 4.74. Found: C, 64.99; H, 5.02. Compound **29**: yield, 73%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  10.37 (s, 1H), 7.52 (s, 1H), 7.35–7.41 (m, 3H), 7.18 (m, 3H), 5.17 (s, 2H), 3.17 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  188.91, 155.03, 153.63, 139.33, 133.50, 130.49, 128.04, 127.40, 126.15, 125.15, 122.96, 115.91, 110.65, 69.74, 55.60 ppm; MS (EI)  $m/z$  276 ( $M^+$ ); Anal. Calcd for  $C_{15}H_{13}O_3Cl$ : C, 65.11; H, 4.74. Found: C, 65.32; H, 4.74. Compound **30**: yield, 63%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  10.36 (s, 1H), 7.39–7.51 (m, 4H), 7.16–7.21 (m, 3H), 5.19 (s, 2H), 3.72 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  188.79, 154.89, 153.38, 135.62, 132.54, 129.37, 128.45, 124.94, 122.88, 115.98, 110.30, 69.63, 55.47 ppm; MS (EI)  $m/z$  276 ( $M^+$ ); Anal. Calcd for  $C_{15}H_{13}O_3Cl$ : C, 65.11; H, 4.74. Found: C, 65.21; H, 4.69. Compound **31**: yield, 35%;

$^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.64 (d,  $J = 7.8$  Hz, 1H), 7.89 (d,  $J = 16.0$  Hz, 1H), 7.31–7.48 (m, 6H), 7.13 (d,  $J = 9.1$  Hz, 1H), 7.01 (dd,  $J = 9.1, 3.0$  Hz, 1H), 6.90 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.16 (s, 2H), 3.74 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.89, 153.39, 151.30, 147.47, 136.81, 129.24, 128.52, 127.94, 127.66, 123.43, 118.83, 114.93, 112.97, 70.41, 55.59 ppm; MS (EI)  $m/z$  268 ( $M^+$ ); Anal. Calcd for  $C_{17}H_{16}O_3$ : C, 76.10; H, 6.01. Found: C, 76.01; H, 6.11. Compound **32**: yield, 55%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.62 (d,  $J = 7.8$  Hz, 1H), 7.87 (d,  $J = 16.0$  Hz, 1H), 7.35–7.61 (m, 4H), 7.31 (d,  $J = 3.0$  Hz, 1H), 7.16 (d,  $J = 9.1$  Hz, 1H), 7.03 (dd,  $J = 9.1, 3.0$  Hz, 1H), 6.88 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.21 (s, 2H), 3.75 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.73, 153.63, 151.02, 147.20, 134.01, 132.73, 130.284, 129.99, 129.46, 129.23, 127.40, 123.65, 118.83, 115.04, 112.81, 68.26, 55.57 ppm; MS (EI)  $m/z$  276 ( $M^+$ ); Anal. Calcd for  $C_{17}H_{15}O_3Cl$ : C, 67.44; H, 4.99. Found: C, 66.99; H, 5.01. Compound **33**: yield, 39%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.65 (d,  $J = 7.8$  Hz, 1H), 7.90 (d,  $J = 16.0$  Hz, 1H), 7.52 (s, 1H), 7.31–7.44 (m, 3H), 7.31 (d,  $J = 2.7$  Hz, 1H), 7.12 (d,  $J = 9.1$  Hz, 1H), 7.03 (dd,  $J = 9.1, 2.7$  Hz, 1H), 6.89 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.19 (s, 2H), 3.74 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.95, 153.54, 151.00, 147.38, 139.44, 133.18, 130.51, 129.33, 127.92, 127.44, 126.27, 123.53, 118.86, 114.97, 112.93, 69.51, 55.61 ppm; MS (EI)  $m/z$  302 ( $M^+$ ); Anal. Calcd for  $C_{17}H_{15}O_3Cl$ : C, 67.44; H, 4.99. Found: C, 66.99; H, 5.01. Compound **34**: yield, 32%;  $^1H$  NMR (200 MHz,  $DMSO-d_6$ ):  $\delta$  9.60 (d,  $J = 7.8$  Hz, 1H), 7.89 (d,  $J = 16.0$  Hz, 1H), 7.42–7.52 (m, 4H), 7.31 (d,  $J = 2.9$  Hz, 1H), 7.13 (d,  $J = 9.1$  Hz, 1H), 7.02 (dd,  $J = 9.1, 2.9$  Hz, 1H), 6.88 (dd,  $J = 16.0, 7.8$  Hz, 1H), 5.18 (s, 2H), 3.74 (s, 3H) ppm;  $^{13}C$  NMR (50 MHz,  $DMSO-d_6$ ):  $\delta$  194.93, 153.50, 151.08, 147.36, 135.88, 132.55, 129.57, 129.28, 128.54, 123.49, 118.85, 114.96, 112.93, 69.58, 55.61 ppm; MS (EI)  $m/z$  302 ( $M^+$ ); Anal. Calcd for  $C_{17}H_{15}O_3Cl$ : C, 67.44; H, 4.99. Found: C, 67.32; H, 5.02.

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